K-ras is currently accepted as the most frequently mutated oncogene in nonsmall cell lung cancer (NSCLC, including squamous carcinoma, adenocarcinoma, and large cell carcinoma). NSCLC patients with the K-ras mutation appear to be refractory to the majority of systemic therapies. In the present study, the in vitro antitumor effects and correlated molecular mechanisms of sorafenib combined with gemcitabine or pemetrexed were explored in the K-ras mutation-positive NSCLC A549 cell line. Sorafenib was seen to exhibit dose-dependent growth inhibition in the A549 cells, while sorafenib combined with pemetrexed demonstrated a greater synergism compared with sorafenib combined with gemcitabine. Sorafenib arrested the cell cycle at the G1 phase, while gemcitabine and pemetrexed caused arrest at the S phase. The molecular mechanism of this synergism was due to the downstream signalling pathways, which were efficiently suppressed by sorafenib, therefore increasing the incidence of the entry of the chemotherapeutic drugs into the apoptotic pathways. Moreover, sorafenib and pemetrexed demonstrated stronger synergism, demonstrating that inhibiting the Ras/Raf/Mek/Erk and Ras/ PI3K/Akt pathways concurrently may achieve improved antitumor effects.

Key words: sorafenib, gemcitabine, pemetrexed, lung cancer, K-ras mutations, western blot.

Contemp Oncol (Pozn) 2016; 20 (1): 33-38 DOI: 10.5114/wo.2016.58499

Synergistic effects of sorafenib in combination with gemcitabine or pemetrexed in lung cancer cell lines with K-ras mutations

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Introduction

Recent data has shown that chemotherapy for non-small cell lung cancer (NSCLC) has reached a therapeutic plateau for overall survival and quality of life [1, 2]. At the same time, the small-molecule epidermal growth factor receptor (EGFR) tyrosine kinase inhibitors (TKIs) that improve the survival of NSCLC patients are causing a paradigm shift in the treatment of NSCLC. Patients with EGFR-activating mutations may greatly benefit from treatment with EGFR TKIs [3–5]. However, K-ras (codon 2 and exon 12) mutations have been correlated with non-responsiveness to EGFR TKIs [6, 7] and systemic chemotherapy [8]. K-ras mutations are found with a high frequency in a variety of human tumours, including 20–40% of NSCLC cases [9]. Thus, clinical research of new treatment strategies for NSCLC patients with K-ras mutations is urgently required.

The Ras/Raf/Mek/Erk and Ras/PI3K/Akt pathways interact with each other to regulate growth, and they play key roles in the transmission of proliferative signals in tumorigenesis. In NSCLC, the activation of K-ras leads to ERK1/2 overexpression through the Raf-Mek-Erk signalling pathway [10-12]. Sorafenib (BAY 43-9006) is an oral multikinase inhibitor that decreases the activity of C-raf and B-raf in the Raf/Mek/Erk signalling pathway and targets the vascular endothelial growth factor receptor (VEGFR-2 and VEGFR-3), platelet-derived growth factor receptor- β (PDGFR- β) families, fibroblast growth factor receptor (FGFR-1), as well as FLT-3, c-KIT, and RET [13]. Sorafenib has been shown to inhibit the growth of a variety of tumour xenograft models driven by the upregulation of Raf/MEK/ERK signalling [14]. Single-agent sorafenib has shown clinical activity in heavily pretreated patients with NSCLC. Stable disease was observed in 30 of 51 (59%) patients treated with 400 mg sorafenib twice daily [15]. In xenograft models, when combining sorafenib with anti-cancer agents, including vinorelbine, cisplatin, and gefitinib, the anti-proliferative effect is at least as effective as sorafenib alone and is well-tolerated [16]. The safety profile of sorafenib in previous trials has increased the feasibility of its use in combination with cytotoxic and cytostatic agents.

Gemcitabine is a pyrimidine nucleoside antimetabolite agent with a favourable toxicity profile, which is active against a variety of human malignancies, particularly NSCLC [17]. Gemcitabine has frequently been used in combinatorial treatments with other anticancer agents.

Pemetrexed is a new antifolate that has significant activity against a broad spectrum of solid tumours, including lung cancer [18, 19]. Pemetrexed inhibits multiple enzymes involved in folate metabolism, including thymidylate synthase, dihydrofolate reductase, and glycinamide ribonucleotide formyltransferase, thereby depleting nucleotide pools and blocking DNA synthesis [20, 21]. In the present study, *in vitro* NSCLC cells harbouring K-ras mutations were used to investigate the effect of sorafenib combined with gemcitabine or pemetrexed on proliferation and cell cycle progression and the molecular mechanisms behind the synergistic effects.

Material and methods

Drugs

Sorafenib (BAY 43-9006) was obtained from Bayer (Leverkusen, Germany) and dissolved in dimethyl sulfoxide (DMSO) to a stock concentration of 10 mmol/l. Gemcitabine and pemetrexed were gifts provided by Eli Lilly that were then dissolved in DMSO at 100 mmol/l as the stock solution. The drugs were stored at -20° C and diluted with culture medium prior to use.

Cell lines

The K-ras mutation human NSCLC A549 cell line was purchased from the American Type Culture Collection (ATCC, Manassas, VA, USA) and maintained in RPMI-1640 medium (Hyclone, Waltham, MA, USA), supplemented with 10% heat-inactivated foetal bovine serum (Hyclone), penicillin (100 U/ml), streptomycin (100 μ g/ml), and L-glutamine (2 mM) at 37°C in 5% CO₂. The cells were then harvested with trypsin-EDTA when they had reached a point of exponential growth.

Anti-proliferative effects of single agents

The anti-proliferative effects of sorafenib, gemcitabine, and pemetrexed as single agents on the A549 cells were evaluated using an MTT assay, as previously described [22]. The cells (4000 cells per well) were cultured in 96-well plates. The IC50 value, indicating the concentration resulting in inhibition of 50% of the maximal cell growth, was determined following 72 hours of exposure to the drugs, as compared with unexposed control cells. Subsequent to the cells being exposed to each drug for 72 hours in 96well plates, 20 µl MTT solution was added to each well. The optical density (OD) of each well was measured at 490 nm following incubation for 4 hours. The percentage of cell growth inhibition resulting from each drug was calculated as: (OD of 490 control cells - OD of 490 treated cells) / OD of 490 control cells × 100. This assay was repeated in more than three independent experiments.

Anti-proliferative effects of sorafenib combined with gemcitabine or pemetrexed

A549 cells were concurrently exposed to sorafenib combined with gemcitabine or pemetrexed for 72 hours. The combination drug doses were selected using constant ratios of the IC50 values. Thus, the combination index (CI) value was calculated using 0.125, 0.25, 0.5, 1, 2, and 4 times the IC50 of the drug combination doses. The CI values were analysed according to the method used by Chou and Talaly using CompuSyn software (ComboSyn, Inc., Paramus, NJ, USA). CI > 1, CI = 1, and CI < 1 indicated antagonistic, additive, and synergistic effects, respective-ly [23].

Cell cycle analysis

The cells (1×10^{5} /well) were plated onto 6-well plates and exposed to sorafenib and gemcitabine (or pemetrexed) as single agents and in combination at the concentration of the IC50 levels for 72 hours. At the end of each exposure, the cells were collected and fixed with 70% cold ethanol at 4°C overnight. DNA staining was performed using a solution with propidium iodide (0.05 mg/ml) and RNase (2 mg/ml) for 30 minutes at room temperature. The cells were analysed using a FACScan cytometer (Becton-Dicknson, Franklin Lakes, NJ, USA), and the percentage of cells in G1, S, and G2 phases of the cell cycle was estimated using Cell Lab Quanta SC software (Beckman Coulter, Miami, FL, USA).

Western blot analysis

The cells (5 × 10⁵/well) were treated with sorafenib and gemcitabine (or pemetrexed) as single agents and in combination at the concentration of the IC50 levels for 72 hours. The cells were washed with ice-cold phosphate-buffered saline (PBS) solution and scraped in lysis buffer. The lysates were centrifuged at 14,000 rpm for 30 minutes at 4°C and the supernatant was collected. Equivalent amounts of protein were analysed by sodium dodecyl sulphate-polyacrylamide gel electrophoresis (SDS-PAGE) and then transferred to PVDF membranes. Appropriate primary antibodies to pAKT, AKT, pERK1/2, ERK1/2, Bcl-2, and β -actin, purchased from Cell Signaling Technology (Beverly, MA, USA), were used. The proteins were visualised with a horseradish peroxidase-coupled secondary antibody from Cell Signaling Technology. Specific bands were detected using an enhanced chemiluminescence reagent (ECL; Perkin-Elmer Life Sciences, Inc., Boston, MA, USA) on autoradiographic film and quantitated by densitometry.

Statistical analysis

The results obtained from at least three independent experiments are expressed as the mean \pm standard deviation. The Student's *t* test and one-way ANOVA test were used to determine the differences between the control and treatment groups. *P* < 0.05 was considered to indicate a statistically significant difference.

Results

Anti-proliferative effects

Sorafenib, gemcitabine, and pemetrexed demonstrated a dose- and time-dependent anti-tumoural effect in the A549 cells when used as single agents (Fig. 1). Subsequent to a 72-hour exposure, the IC50 values of sorafenib, gemcitabine, and pemetrexed in the A549 cells were 4.09 \pm 0.16, 11.54 \pm 0.77, and 3.90 \pm 0.40 μ M, respectively.

When exposed to the concurrent administration of sorafenib and gemcitabine, synergistic activity (mean CI value, 0.86) was observed in the A549 cells. Moreover, the concurrent administration of sorafenib and pemetrexed generated a greater synergistic effect (mean CI value, 0.63; Figs. 2 and 3).

Cell cycle effects

Flow cytometry was applied to evaluate the cell cycle phase distributions in the A549 cells following single-drug and concurrent administration of sorafenib and gemcitabine (or pemetrexed) for 72 hours. Following sorafenib treatment, the proportion of GO/G1 phase cells increased relative to the control values (p < 0.01). With gemcitabine or pemetrexed treatment alone, the fraction of S phase cells increased (p < 0.01).

Upon exposure to a concurrent administration of sorafenib and gemcitabine, the proportion of GO/G1 phase and S-phase cells increased compared with the control (p < 0.05). However, when exposed to sorafenib combined with pemetrexed, the proportion of GO/G1 phase cells decreased, while the proportion of G2/M phase cells markedly increased (Fig. 4).

Effects on downstream signalling pathways

Following a 72-hour exposure at the IC50 concentration, gemcitabine was demonstrated to upregulate the levels of p-ERK, p-AKT, and Bcl-2, whereas pemetrexed upregulated the levels of p-ERK and Bcl-2, but decreased the level of p-AKT, and sorafenib downregulated the levels of p-ERK, p-AKT, and Bcl-2, compared with the control A549 cells.

In addition, when exposed to a concurrent administration of sorafenib and gemcitabine, the levels of p-ERK and Bcl-2 were downregulated, while the level of p-AKT remained identical to that of the control group. When exposed to sorafenib combined with pemetrexed for 72 hours, the levels of p-ERK and Bcl-2 decreased and the level of p-AKT further declined (Fig. 5).

Discussion

K-ras is currently accepted to be the most frequently mutated oncogene in NSCLC. It represents the activating mutation in the Ras/Raf/Mek/Erk signalling pathway. Sorafenib is a novel, multikinase inhibitor that targets tumour proliferation and angiogenesis [13]. The present study was performed on the K-ras mutation A549 human lung cancer cell line to investigate the anti-proliferative effects of sorafenib as a single agent and in combination with chemotherapy using gemcitabine or pemetrexed.

Sorafenib, gemcitabine, and pemetrexed were demonstrated to exhibit dose-dependent growth inhibition of cell growth in the A549 cells when used as single agents. The combination of sorafenib with gemcitabine or pemetrexed generated synergistic effects in the A549 cells. Moreover, greater synergism was observed upon use of sorafenib combined with pemetrexed.

In the present study, the A549 cells were arrested at the G1 phase by sorafenib, while gemcitabine and pemetrexed caused S-phase cell accumulation. When exposed to a concurrent administration of sorafenib and gemcitabine, the proportion of G0/G1 phase and S-phase cells increased. These effects on the cell cycle may explain the synergism observed between the two drugs, since the two pharmacological agents did not antagonise their mutual mechanism of action.



Fig. 1. *In vitro* effects of sorafenib, gemcitabine, and pemetrexed on proliferation of A549 cell lines. MTT assays were used to examine the inhibitory activities on cell proliferation. Each data point was repeated in more than three independent experiments

When exposed to sorafenib combined with pemetrexed, the proportion of GO/G1 phase cells was significantly reduced, while the proportion of G2/M phase cells was significantly increased. This may be due to the fact that the cells in the G1 phase entered into the apoptotic pathway and died, so the proportion in the G2/M phase was relatively increased.

The present study also found that gemcitabine enhanced the expression levels of certain downstream sig-



Fig. 2. A549 cells were exposed to different combinations at IC50 levels. (OLYMPUS IX70 ×200 magnification). The anti-proliferative effects of sorafenib combined with pemetrexed were more pronounced than sorafenib combined with gemcitabine





Fig. 3. The combination index (CI) value of the two combinations was calculated using the Chou-Talalay method. S+G means sorafenib and gemcitabine, S+P means sorafenib and pemetrexed. CI < 1 was detected in both of the two combinations; sorafenib and pemetrexed generated stronger synergistic effects

Fig. 4. Flow cytometric analysis was applied to determine the alterations in cell cycle distributions in A549 cell lines. The concentrations of sorafenib, gemcitabine, and pemetrexed were used at IC50 levels. Columns in the diagram depict cell cycle phase distribution in A549 cells



Fig. 5. The expression levels of downstream signalling pathways in A549 cells were evaluated by western blot analysis

nalling pathways, increasing the levels of p-AKT, p-ERK, and Bcl-2 in the A549 cells. Pemetrexed also enhanced the expression levels of p-ERK and Bcl-2. In accordance with these results, a previous study reported that the cell signalling pathways may be gradually activated by chemotherapy [24]. p-ERK and p-AKT play significant roles in tumour cell proliferation; chemotherapy increases the levels of p-ERK and p-AKT, leading to the prevention of apoptosis. We also found that sorafenib may downregulate the levels of downstream p-AKT, p-ERK, and Bcl-2 in the A549 cell lines following 72 hours of exposure.

When sorafenib was combined with gemcitabine or pemetrexed, a significant decrease was observed in the p-ERK and Bcl-2 levels in the A549 cells compared with the control cells. We conclude that the RAF/MEK/ERK signalling pathways activated by gemcitabine or pemetrexed may be blocked by sorafenib, thus increasing the incidence of the entry of the chemotherapeutic drugs into the apoptotic pathways. These observations on p-ERK and Bcl-2 in NSCLC cells may explain the synergistic growth inhibitory effects of sorafenib combined with gemcitabine or pemetrexed.

In the present study, pemetrexed and sorafenib downregulated the level of p-AKT individually, and when the two drugs were combined the level of p-AKT was further decreased. When using a combination of sorafenib and gemcitabine, the level of p-AKT was neither increased by the gemcitabine nor decreased by the sorafenib, but remained identical to that of the control group. Thus, this is why the combination of sorafenib and pemetrexed generates a stronger synergistic effect than sorafenib and gemcitabine. The simultaneous inhibition of p-AKT and p-ERK generates greater anti-proliferative effects.

In conclusion, we found that sorafenib caused significant growth inhibition in the NSCLC cells with K-ras mutation. Moreover, sorafenib combined with pemetrexed generated greater synergistic effects than sorafenib and gemcitabine against the NSCLC cells with K-ras mutation. These data are encouraging for the development of sorafenib as a single targeted therapy or in combination with cytotoxic chemotherapy drugs for the treatment of NSCLC cells with K-ras mutation. Further studies are needed to explore *in vivo* sorafenib as a single agent and combined with gemcitabine or pemetrexed in lung cancer cell lines with K-ras mutations.

The authors declare no conflict of interest.

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Submitted:28.10.2013Accepted:11.06.2014